

APPLICATION NOTE

Achieve unmatched high-throughput sample analysis with the Aurora Rapid 8x150

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Aurora Rapid 8x150 packed emitter column for CaptiveSpray (8 cm x 150µm ID, 1.7µm C18) Part No. AUR3-80150C18A-CS1



Aurora Rapid 8x150 packed emitter column for EASY-Spray and Nanospray Flex (8 cm x 150µm ID, 1.7µm C18) Part No. AUR3-80150C18A-TS

INTRODUCTION

Robust performance across a large patient plasma cohort analysis

IonOpticks' Aurora Rapid 8x150 packed emitter columns feature our revolutionary nanoZero® fitting, allowing for simplified plug-and-play high-throughput proteomics. These columns are capable of analyzing from 50 to more than 100 samples per day while delivering class-leading peptide and protein identifications.

The Aurora Rapid 8x150 columns were used to analyse a large patient plasma cohort on an Evosep One and Thermo Fisher Orbitrap Astral. Weekly quality control (QC) of the LC-MS performance and reproducibility was performed by injection of 200ng HeLa tryptic peptides using a 60 SPD method. Plasma samples were analysed using a 100 SPD method with approximately 700 plasma samples run between QC injections.



High-throughput analysis of peptides and proteins:

HeLa cell tryptic peptides, used for LC-MS QC, were injected and separated using a 60 samples per day method on an Evosep One. This method repeatedly identified over 9,300 proteins per run, with an average of 100,000 unique peptide identifications.

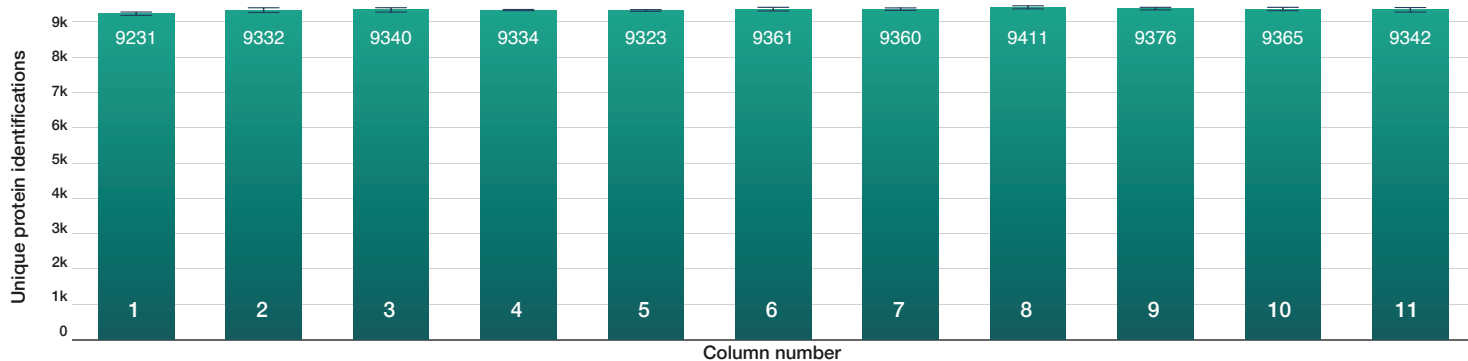


Figure 1: Protein identifications across different columns. A HeLa tryptic digest (200 ng) was separated on an Aurora Rapid 8 cm x 150 µm column on a 60SPD method using Evosep One and Thermo Fisher Orbitrap Astral Mass Spectrometer.

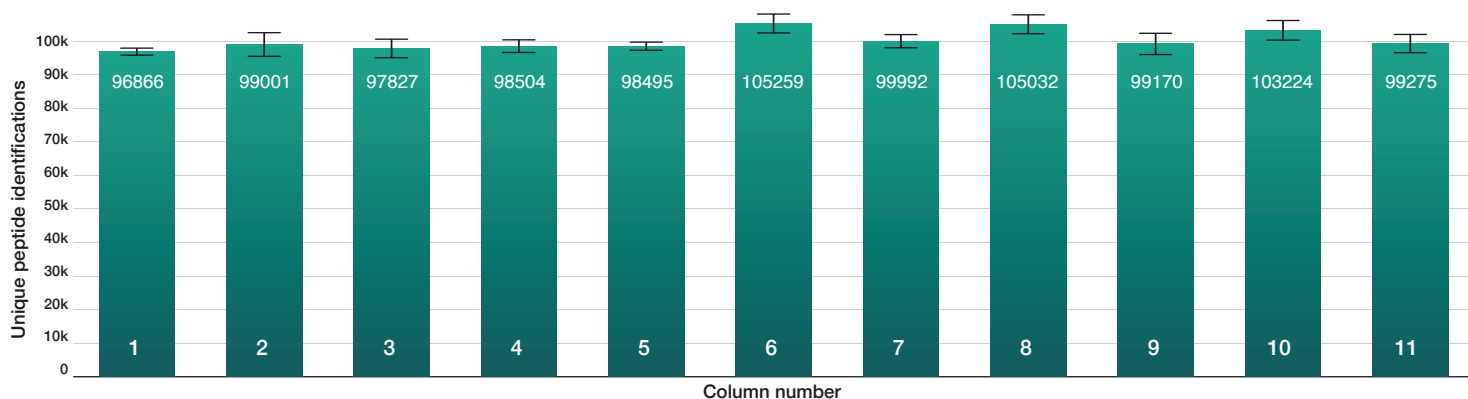


Figure 2: Unique peptide identifications across different columns. A HeLa tryptic digest (200 ng) was separated on an Aurora Rapid 8 cm x 150 µm column on a 60SPD method using Evosep One and Thermo Fisher Orbitrap Astral Mass Spectrometer.

High reproducibility across multiple columns

To assess the reproducibility of the protein quantification across multiple QC runs and columns, protein intensities were used to calculate a coefficient of variation (CV) and Pearson correlation matrix, revealing uniform performance with low CV values. Pearson correlation matrix visualization further supported this, showing high correlations between runs (mean Pearson correlation coefficient: 0.98).

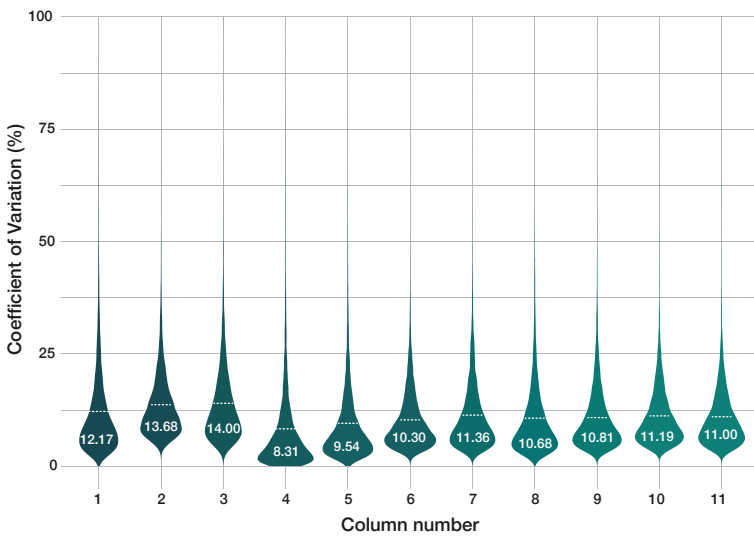


Figure 3: Low CVs across different columns: Violin plot of coefficient of variation for all identified protein intensities from HeLa tryptic digest QC injections on Aurora Rapid 8 cm x 150 μm columns. Samples were run on an Evosep One and Thermo Fisher Orbitrap Astral Mass Spectrometer. Dashed line represents the median value for each column.



Figure 4: A Pearson correlation matrix of all quantified protein intensities from the 257 QC injections was calculated.

Narrow peak widths

The below plot displays the full width at half maximum (FWHM) values across 257 QC runs, revealing consistently narrow peak widths. This consistent performance indicates good column quality and reproducibility, important for large patient cohort analyses.

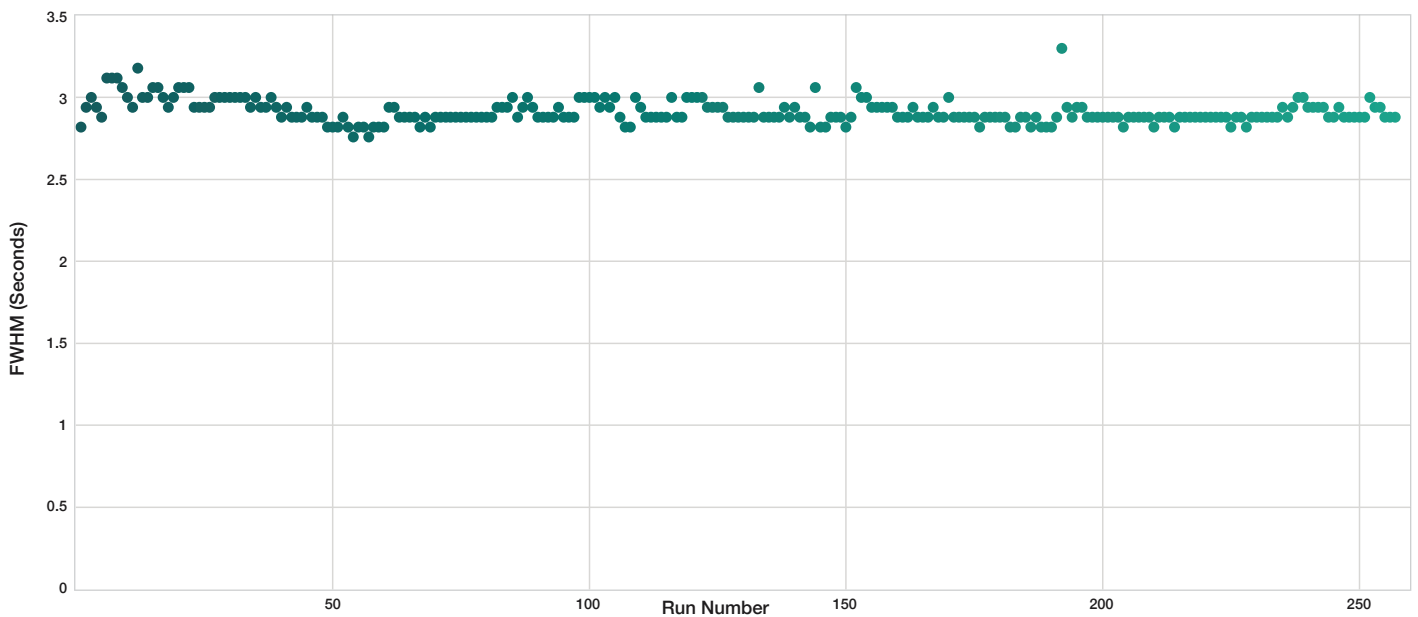


Figure 5: Peak widths across all runs: average full width at half maximum (FWHM) for all identified peptides from HeLa Tryptic Digest injections on an Aurora Rapid 8 cm x 150 μm columns. Samples were run on a Evosep One and Thermo Fisher Orbitrap Astral Mass Spectrometer.

More than

100k

unique peptides
from 200 ng



>0.98

mean Pearson
correlation



2.9

secs
full width
half maximum



Across

257

QC runs

Stable retention times ensure confidence in results

The analysis of retention time reproducibility is detailed through a scatter plot and histogram, focusing on the coefficient of variation for precursor retention times. The scatter plot demonstrates a high level of consistency, with most data points clustering below a CV of 6% and a median CV at 2.52%, showing stable retention times across precursors. Additionally, a four-month experiment on 16 selected peptides showed stable average retention times, further proving the reliability and precision of IonOpticks columns.

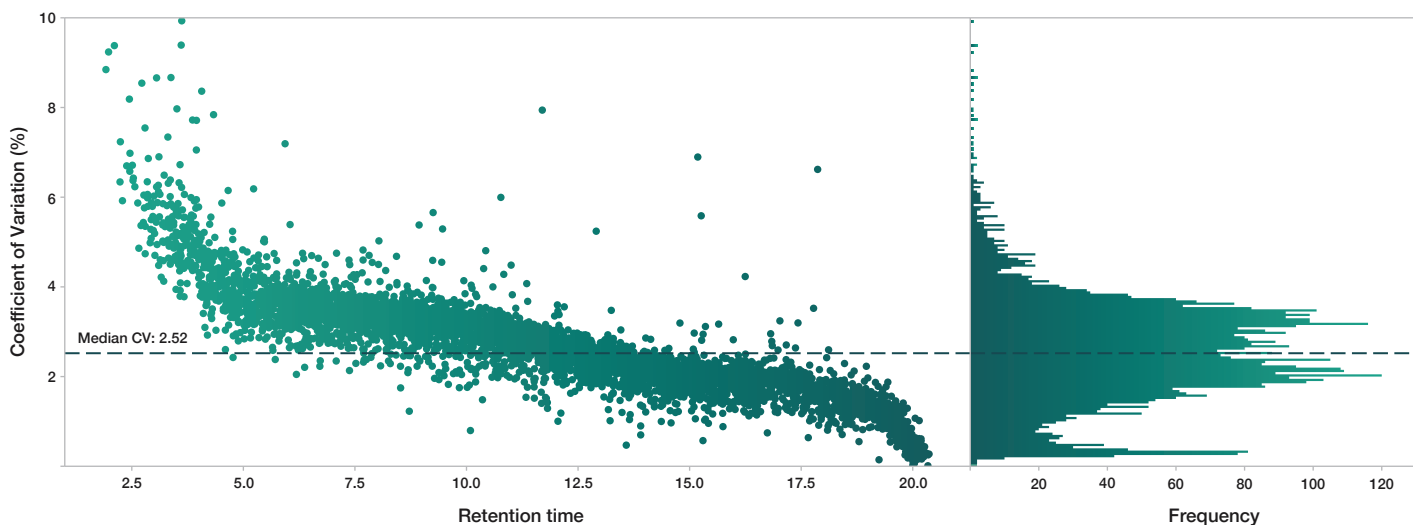


Figure 6: Scatter Plot and histogram analyzing the coefficient of variation of retention times for precursors: each dot represents an individual precursor across all of the 257 HeLa QC runs. The plot shows that most data points cluster below a CV of 6%, with the median CV marked at 2.52, indicating consistent retention times across all precursors.

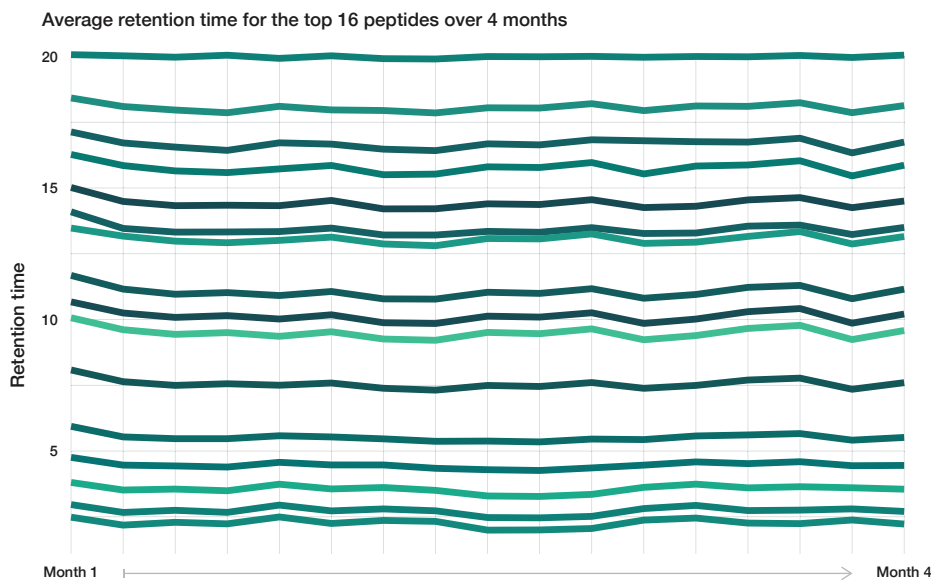


Figure 6: Stable peptide average retention times across 4 month period: 16 peptides were selected and their average retention time assessed across 4 months period from HeLa tryptic digest injections on Aurora Rapid 8 cm x 150 µm columns. Samples were run on an Evosep One and Thermo Fisher Orbitrap Astral Mass Spectrometer.

METHODS

LC-MS Analysis

A HeLa tryptic digest (200 ng) was separated on Aurora Rapid 8 cm x 150 µm column on a 60SPD method using Evosep One and Thermo Fisher Orbitrap Astral Mass Spectrometer.

MS parameters: mass range of MS(Orbi)/MS2(Astral) from 380 to 980, isolation windows of 2Th, a maximum injection time (IT) of 3 ms, FAIMS compensation voltage (CV) set at -40, and a resolution for MS1 set at 120,000.

Data Processing

Data was processed using DIA-NN software (v1.8.1) with match-between-runs enabled. The pg.matrix.tsv, stats.tsv and pr.matrix.tsv tables were used to calculate protein and peptide identifications and metrics.

CONCLUSION

The combination of the IonOpticks Aurora Rapid 8x150 column, the Evosep One and Thermo Fisher Orbitrap Astral Mass Spectrometer enabled maximum protein and peptide identifications whilst also ensuring robust and reproducible protein quantification across a large patient plasma cohort analysis.

ACKNOWLEDGEMENTS

We thank Professor Matthias Mann and his team at the Max Planck Institute of Biochemistry, where all data was acquired and processed. Special thanks to Dr. Igor Paron for his assistance with the LC-MS setup and operation.

ABOUT IONOPTICKS

IonOpticks produces high-performance chromatography solutions for the global research community. We specialise in the development and manufacture of columns for analytical applications in liquid chromatography with mass spectrometry (LC-MS) and high-end proteomics. Our highly reproducible methods provide a unique ability to enhance the sensitivity of mass spectrometry sample analysis, enabling scientists and clinicians to discover more from their samples. Our team are experts in a broad array of LC-MS platform technologies and are driven by the need to improve chromatographic performance in order to achieve data quality and deep proteome coverage on a whole new scale.

FURTHER READING

For further resources and technical support, visit our Help Centre at helpcentre.ionopticks.com. To view other application notes, read the latest publications featuring Aurora Series columns, or view the full range of IonOpticks products, visit our website at www.ionopticks.com